

Hydrogen evolution reaction: Optimizing a didactically reconstructed system with variable control strategies

Malte Petersen[†], Jonas Eichhorn[†], Felix H. Schacher, Timm Wilke^{*}

Introduction

The generation of hydrogen with sunlight is becoming increasingly important and is a central building block for a sustainable energy supply. One approach in current research is to embed a light-driven catalytic molecule within a hierarchically structured soft matter matrix to use solar radiation within a chemical reaction for photocatalytic water splitting. In previous work, a first proposal for the didactic reconstruction of this research area was already presented (Fig. 1) [1]. Besides the high didactic potential of current cutting-edge scientific research on sustainability itself, the experiments can be used to familiarize students with basic scientific research methods.

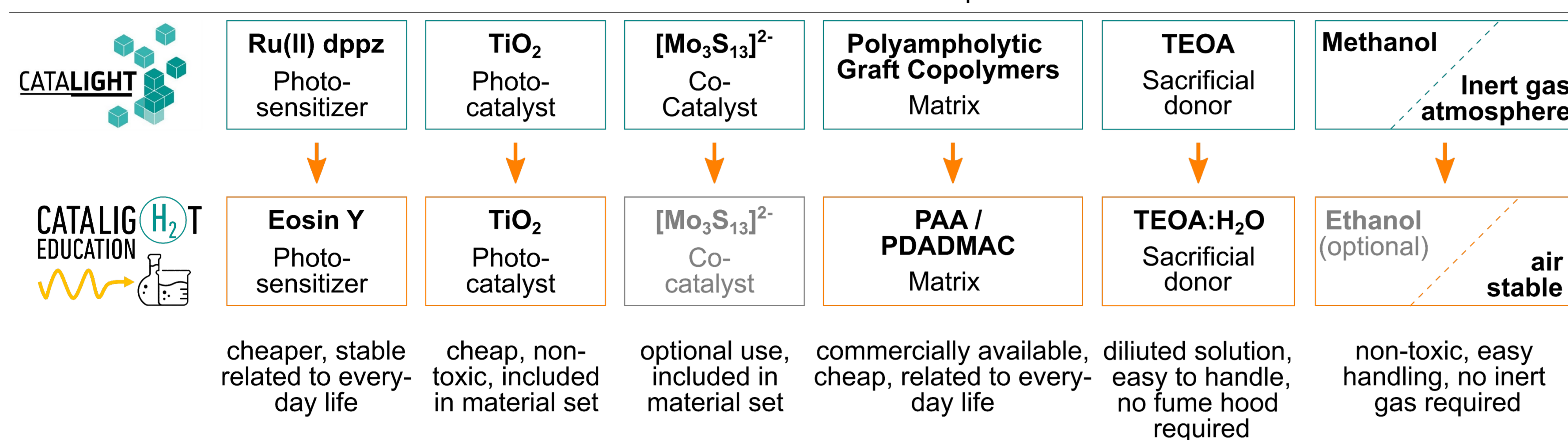


Fig. 1: Development of a simple system for photocatalytic water splitting for student laboratories. The changes compared to the system which is used for CATALIGHT research are marked in orange.

Scientific Background

The photocatalytically active system consists of three essential components. The **catalyst** TiO₂ (CAT) nanoparticles alone are able to perform photocatalytic hydrogen evolution upon UV-light irradiation acting as light harvesting unit and catalyst at the same time [2]. To shift needed light energy into the visible region broadening the use of the solar spectrum the usage of a suitable **photosensitizer** eosin Y (PS) provides TiO₂ with these necessary electrons. Thus, the excited PS enables the transfer of electrons from the **sacrificial electron donor** triethanolamine (TEOA), provides electrons needed for reducing protons to molecular hydrogen, onto the CAT, which in turn is able to reduce protons to molecular hydrogen (Fig. 2).

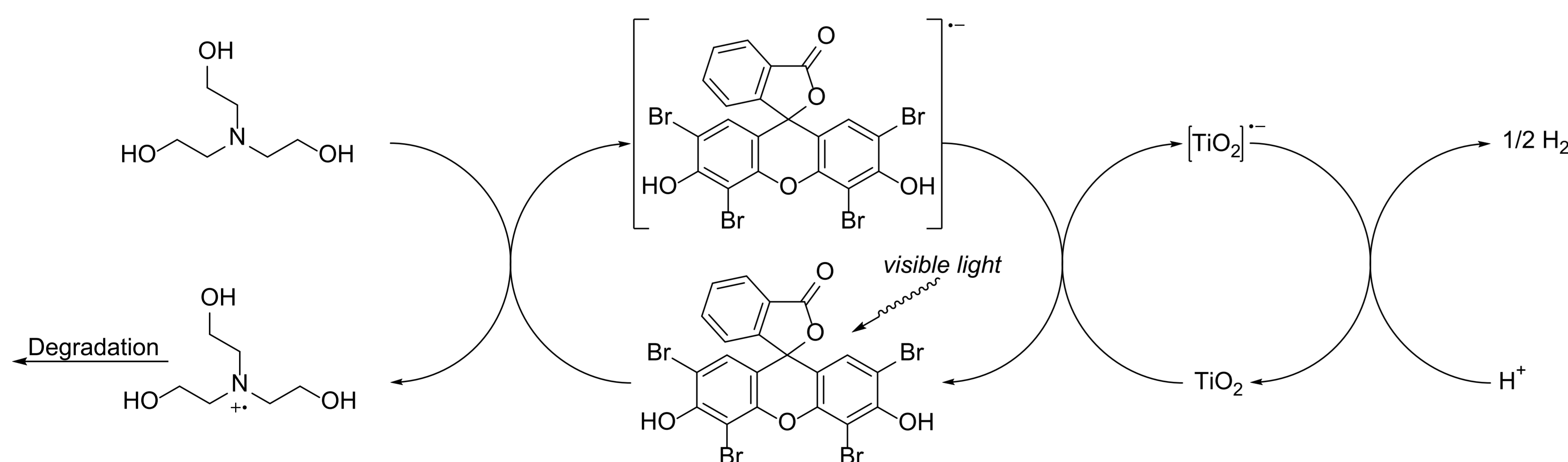


Fig. 2: Reaction scheme for visible light-induced hydrogen evolution catalysis using TEOA as sacrificial electron donor, eosin Y as photosensitizer and TiO₂ as catalyst.

In this work a combination of commercially available polymers forming polyampholytic complexes is used to mimic the unique stabilizing properties of a tailor-made graft-copolymer from the literature. In this context, numerous reports exploit the ability of poly(acrylic acid) ("PAA") and poly(diallyldimethylammonium chloride) ("PDADMAC") to form pH and ionic strength dependently complexes in solution [3]. With this polyelectrolyte combination we target a suitable soft matter matrix to perform photocatalytic investigations.

Control-of-variables Strategy

The common starting point is the observation that experiments in the classroom are often characterized by the central objective of imparting knowledge. In the process, independent experimentation is pushed into the background by the provision of detailed experimental instructions [4]. By enabling different decisions for the further planning of an experiment, the students can independently experience and acquire the central feature of scientific working methods, the variation of a single variable with simultaneous constancy of the other components. Only through this experimental exploration of the individual components and their different influences is it possible for them to establish a cause-effect relationship.

Experiment & Results

In the presented example, the influence of different concentration changes of the polymermatrix on the degradation rate of PS is to be investigated. The corresponding time dependent UV/vis spectra are shown in Fig. 4. The assumption is that with an accelerated degradation the reactivity of eosin Y is increased. The acceleration of the degradation of eosin Y while increasing the polymer concentration, most prominently visible upon comparing the absorption spectrum before and after irradiating for 20 min, can be attributed to an improved dispersing effect and increasing electrostatic interactions of the components introduced by the polymers used.

References

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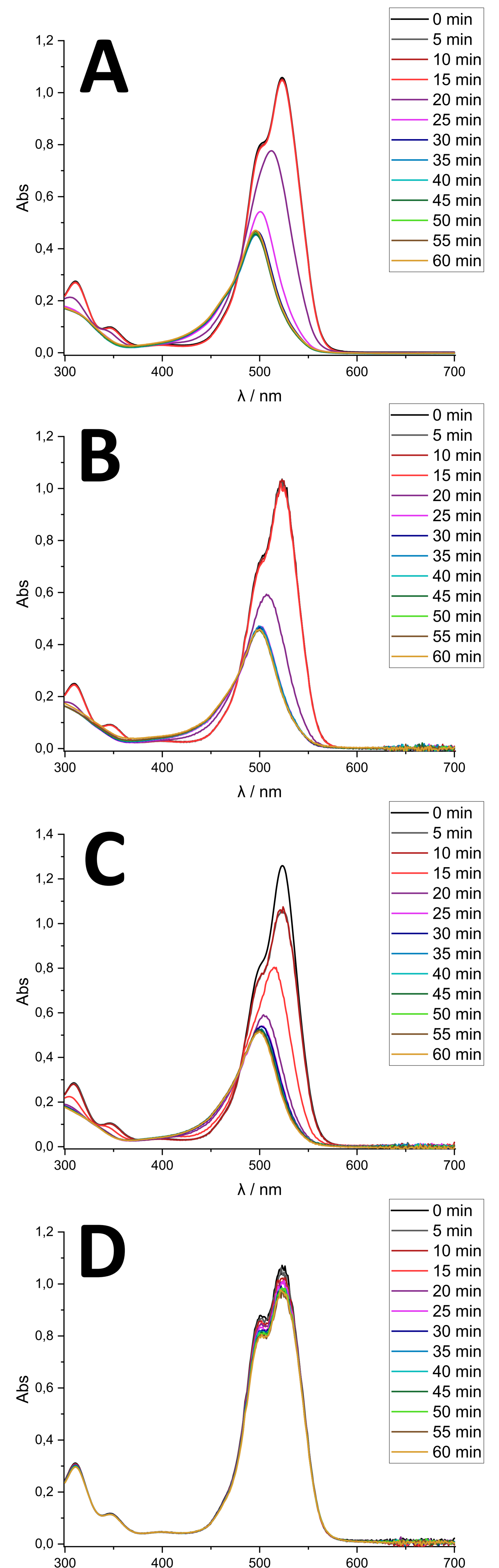


Fig. 4: UV/vis spectra of different kinetic investigations, c(eosin Y) = 22.2 μM, c(TiO₂) = 83.4 μM, c(TEOA) = 83.3 mM. (A): Sample A (c(PAA) = 0.17 μM, c(PDADMAC) = 0.23 μM), (B): Sample B (c(PAA) = 0.34 μM, c(PDADMAC) = 0.46 μM), (C): Sample C (c(PAA) = c(PAA) = 0.51 μM, c(PDADMAC) = 0.69 μM), (D): Sample A without TEOA.



Malte Petersen
 Research Assistant
 Friedrich Schiller University Jena
 Chemistry Education Department
 August-Bebel-Str. 2, D-07743 Jena
 m.petersen@uni-jena.de

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